

# European Bank for Induced Stem Cells

The EBISC - European Bank for induced pluripotent Stem Cells project has received support from the Innovative Medicines Initiative Joint Undertaking under grant agreement n° 115582, resources of which are composed of financial contribution from the European Union's Seventh Framework Programme (FP7/2007-2013) and EFPIA companies' in kind contribution. [www.imi.europa.eu](http://www.imi.europa.eu)

# Today's lesson.....



- 
- EBiSC
    - What is it?
    - What has been done so far?
  - Industrialisation of hiPSC
    - The things we need to remember.
    - The things we need to do (exemplar).

# Mission



## Standardise hiPSC

- Procurement
- Banking
- Quality Control
- Distribution

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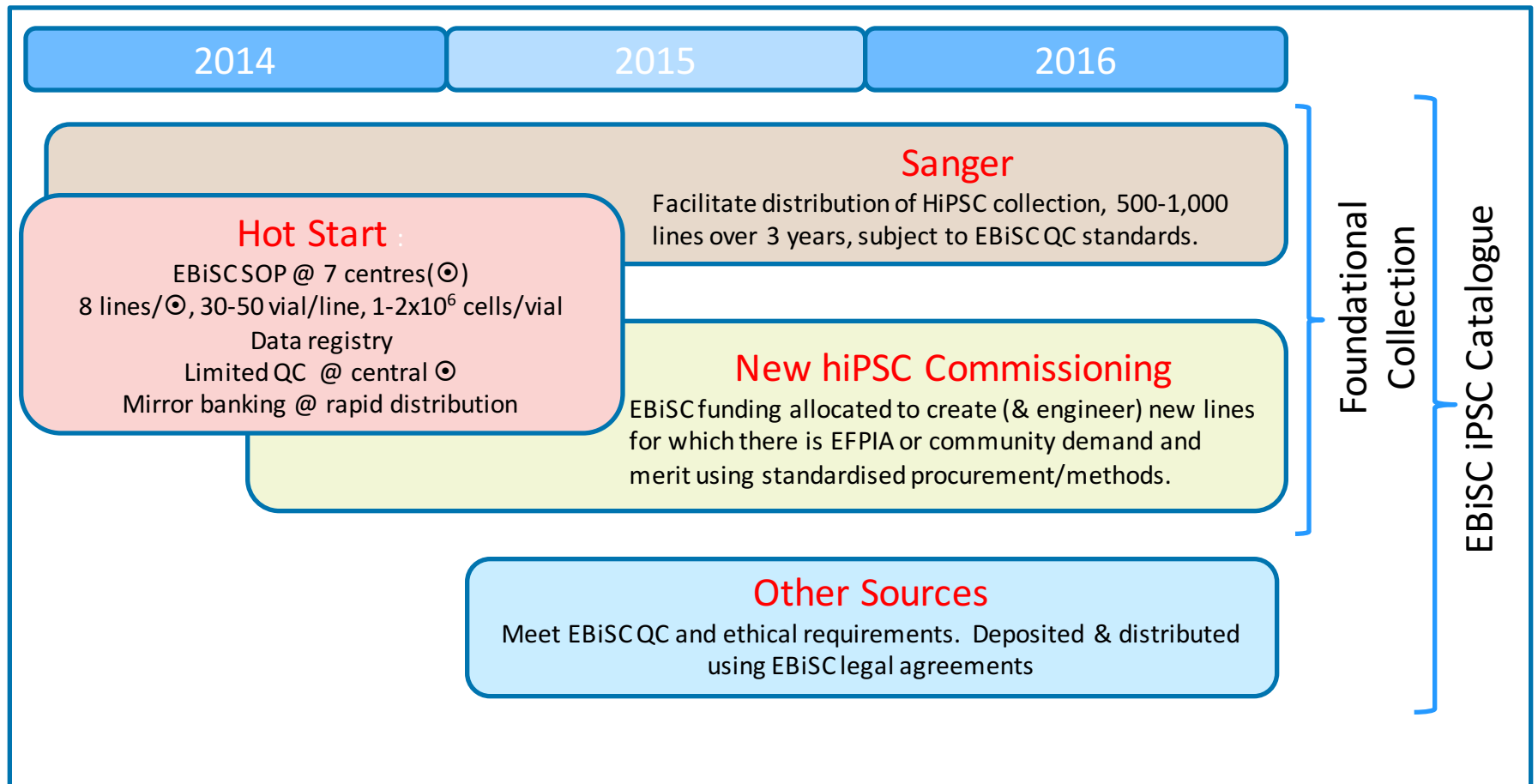
- Pfizer co-ordinates
- Roslin Cells Ltd manages
- Janssen Pharmaceutica NV
- UCB Biopharma SPRL
- H. Lundbeck A/S
- Novo Nordisk A/S
- AstraZeneca AB
- Bayer
- University of Edinburgh
- Charite University Medicine Berlin
- University of Newcastle Upon Tyne
- Klinikum Der Universitaet Zu Koeln
- The Hubrecht Institute
- University College London
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- Fraunhofer
- EMBL – EBI
- Dept. Health
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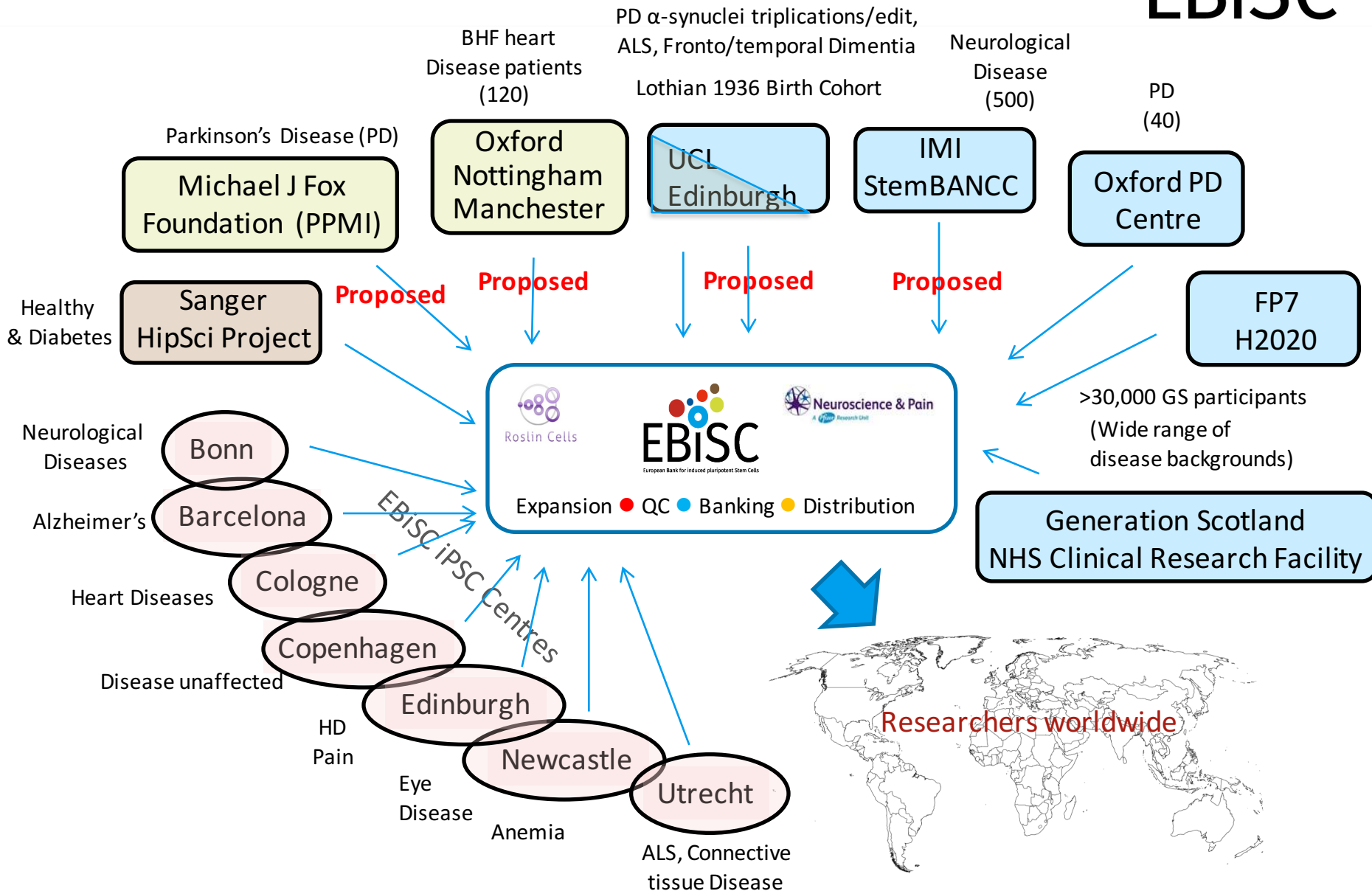
CO – Dr Tim Allsopp



# Establishment of a Foundational Collection & Catalogue



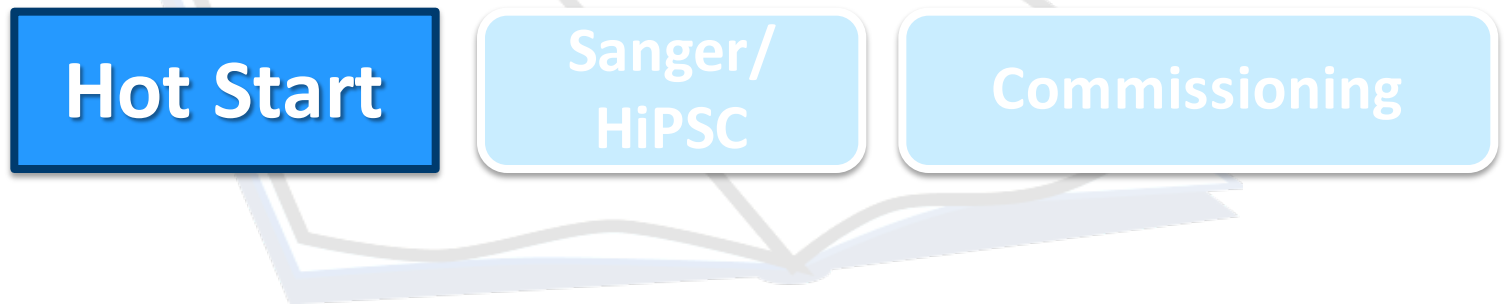
# Disease Coverage & Catalogue Expansion to Date



# Foundational collection: Why Hot Start ?



## The EBiSC Catalogue:



- Getting “wheels on” → best practice via experience
- Procurement
  - No commercial restrictions.
  - Ethical principles.
- Centre characterisation data (hESC-Reg)
- Standardised expansion/cryopreservation/qualification.



# The Hot Start Process – *to get the wheels on*



Clinic

At Derivation Centres

**(Procurement) (Derivation) Expansion**

**Acceptance criterion & SOPs agreed/issued**

- PI&C check
- 1<sup>o</sup> tissue processing
- Donor data capture (Traceable anonymity)

- SOPs reinforced with training
- Feeder free
- Vitronectin or Geltrex
- E8 or mTESR1
- Passaging with EDTA

**Banking**

**Guidance Documents**

- Cryoprotectant
- At harvest cells should be pooled
- Vials should be filled in numerical order
- Labels provided from RC

**Cell line data**

**Forms**

- Original data form
- General cell line information
- Banking and testing form
- Batch specific QC information

UKKi012-A  
14/10/002/P001/001  
NP0041-17  
EBISC  
vial 001  
SAMEA2825982



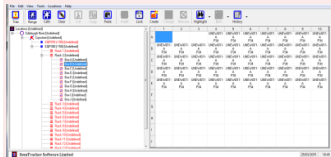
# The Hot Start Process

## At EBiSC Central

### Cell line receipt

Cell Line Registered into cryostorage

EBiSC/SOP/1  
EBiSC/SOP/2  
SOP/EQP/75  
EBiSC/FRM/1



### Thawing of cell line

Thaw into 2 wells of a 6-well plate

EBiSC/SOP/10  
EBiSC/FRM/3



- Viability
- Genetic Identity
- Genetic integrity
- Morphology

### Culture

Passage x3 in antibiotic free media

EBiSC/SOP/9  
EBiSC/FRM/4  
EBiSC/FRM/6



- Sterility
- Mycoplasma
- Virology
- Microorg. Growth
- Pluripotency

### Limited QC + data from ☉

Collate all data and prepare CoA

EBiSC/FRM/14

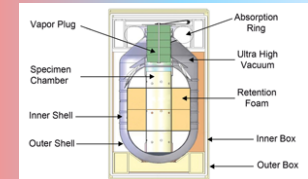
Certificate of Analysis for iPSC			
EBiSC Catalogue No.	44182024	Batch no.	1001
EBiSC Reg / EBiSC cell line name	UMBRO14	Row ID	18-1102-2016
Sex	Male	Year of origin	2016-November
Disease Association	Myofibrilopathy	Phenotype of Disease	Affected
Reprogramming Method	Integrating retrovirus (PGCFL1, SOX2, KLF4, MYC)		
Passage no.	10	Cell number at passage	1.1 x 10 <sup>6</sup>
Culture and passaging methods	Reprogrammed into cells in a well of a 6-well plate. Culture using iPSCgibco medium with 4000i supplement, passaging using 100% feeder cell free protocols for further passage.		
Additional comments	None growth after thaw. High rate of cell growth cycle thereafter.		
Associated Publications	Published in: [2013] [1816]		
The following standard testing criteria have been determined. Like EBiSC, prior to release of this product.			
Test	Assay	Result	
Confirmed Identity	Visual assessment for micro-biological growth	Not Detected	
	Inspection for mycoplasma growth	EBiSC	
Cell Line Identity	qPCR for Mycoplasma	Not Detected	
Cell Line Identity	Strain (2013, 2014, 2015, 2016)	Not Detected	
Cell Line Identity	Short Tandem Repeat analysis using STR	Matched	
Viability post thaw	Ability to re-plate and grow in 6-well	Successful	
Morphology	Continuous visual assessment of iPSC colonies (morphology)	Typical iPSC colonies	
Differentiation Potential	EBiSC differentiation and qPCR for pluripotency markers	Successful	

## Shipment

### Transfer of cells

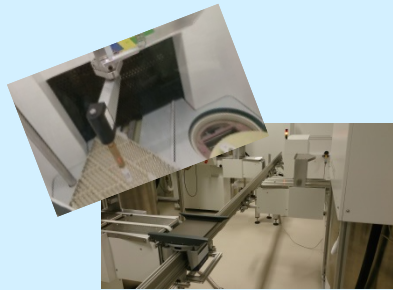
Ship banks to final storage using dry shippers

Hold 2 vials at RC as backup



# The Hot Start Process

## Final Storage



**Fraunhofer**  
IBMT

Mirror bank at  
Fraunhofer (IMBT)  
~10 % of bank



**ECACC**  
European Collection  
of Cell Cultures  
Operated by Public Health England

Final stage  
distribution at  
ECACC  
~85 % of bank

## Customer

Customers access EBISC  
catalogue via IMS & ECACC

Stem Cell Biologists worldwide

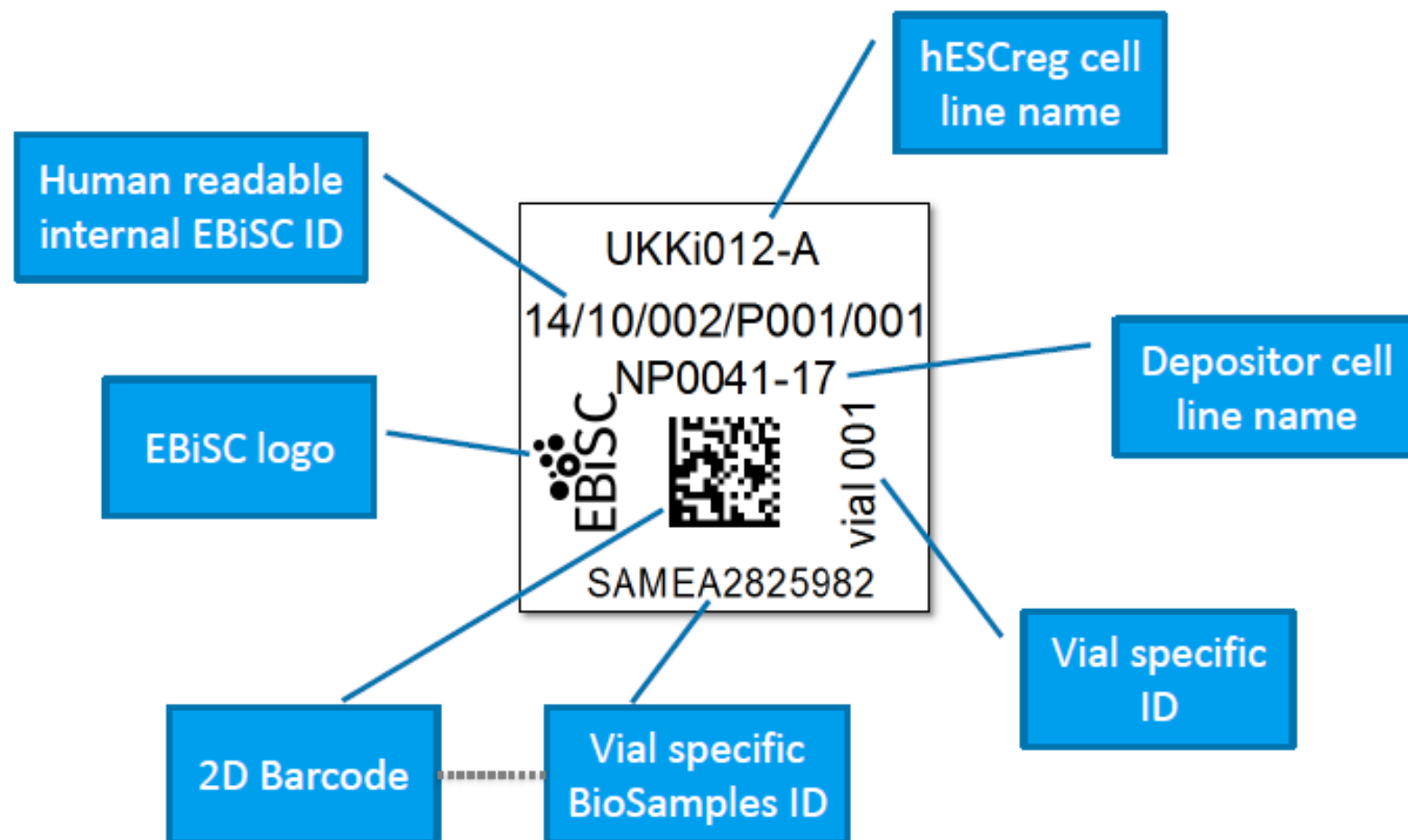


© 2009 www.free-world-maps.com





# Data management – labelling & asset tracking



# How do customers benefit?

## Cell Line & Data Assets



- Vial of cells
- Access and Use Agreement
- Terms and conditions
- Instructions for use
- Culture manuals, SOPs
- Certificates of analysis
- Clarification on QC applied
- Managed access to data
- Customer feedback system

The image shows three overlapping certificates of analysis (COAs) for induced pluripotent stem (iPS) cell lines. The topmost certificate is for cell line UNIK007-A, with ECACC Catalogue No. E6540001. It provides detailed information about the cell line's origin, reprogramming method, and quality control results.

Certificate of Analysis for Induced Pluripotent Stem Cell Line			
ECACC Catalogue No.	E6540001	Batch no.	P001
HESC reg / EBISC cell line name	UNIK007-A	Donor ID	NP0024
Sex	Female	Tissue of origin	Dermal Fibroblasts
Disease Association	Heart Conduction Disease	Phenotype of Donor	Affected
Reprogramming Method	Integrating retrovirus (POU5F1, KLF4, SOX2)		
Passage no.	26	Cell number / vial	1-2 x 10 <sup>6</sup>
Culture and passaging methods.	Recommended thaw into 1 well of a 6-well plate. Culture using Vitronectin with Essential EB medium, passaged using EDTA. Refer to cell line user protocols for further guidance.		
Additional Comments	Typical growth after thaw, typical growth cycle.		
Associated Publications	PubMed ID: 22178870		
The following standard testing criteria have been determined within EBISC, prior to release of this batch:			
Test	Assay	Result	
Sterility	Breth Inoculation for microbiological growth	No Bacterial or Fungal growth detected	
	qPCR for Mycoplasma	Not Detected	
	Virology (HBN, HCV, HIV1, HIV2)	Not Detected	
Cell Line Identity	Short Tandem Repeat analysis (STR)	Recorded	

# The Hot Start – State of Play

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## Number of Centres that have:

Specified contribution

Sent lines to EBiSC central (RC)

Completed Depositor forms

Completed Banking & Testing forms

Supplied P&IC information

Signed EBiSC MDA

**All 8 lines    3-7 lines    0-2 lines**

6                      1                      0

4                      3                      0

4                      2                      1

4                      3                      0

1                      3                      4

3                      0                      4



# Culturing Cell Lines at Roslin Cells: Our experience



## Performance of 47 Hot Start cell lines processed so far

<b>Recovery after thaw</b>	<b>Recovered well</b>	<b>Were difficult to recover</b>	<b>Were not viable</b>
	41	5	1
<b>Cell morphology</b>	<b>Very good iPS morphology</b>	<b>Good iPS morphology</b>	<b>Poor iPS morphology</b>
	26	14	7
<b>Levels of differentiation in culture</b>	<b>Low to Medium</b>	<b>High differentiation</b>	
	40	7	
<b>Sterility issues</b>	<b>Had no issues</b>	<b>That were contaminated</b>	
	44	3	
<b>Cell identity issue</b>	<b>Had no issues</b>	<b>Wrong identity</b>	
	40	7	

# The Hot Start – Lessons learned

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- Depositor concerns re: competitor access.
- ✓ Feeder-free culture conditions.
- Training → SOP compliance.
- ✓ Biosample ID label.
- Interface with hESCreg and EBI.
- ✓ Laboratory Information Management System.
- Member state restrictions on hiPSC distribution.

# The Hot Start - EBiSC Control Lines

<b>hESC reg</b>	UKKi012-A	UKBi005-A
<b>Local cell line ID</b>	NP0041-17	LB-31-rl
<b>Derivation centre</b>	Cologne	Bonn
<b>Disease / Control</b>	Healthy Control	Healthy Control
<b>Gender</b>	Male	Female
<b>Reprogramming method</b>	Episomal	Retroviral
<b>No. vials currently banked.</b>	30	70
<b>HotStart</b>	Initiated 24-Feb-15. EBiSC QC in process.	Expanded bank completed. EBiSC QC in process.
<b>Derivation centre QC</b>	Submitted, some aspects still in progress.	Data in hESCreg
<b>Comments</b>	Initial HotStart shows typical morphology and growth.	Female control line, previously requested by WP6.2.

Well characterised lines for use within the EBiSC consortium to establish consortium-wide SOPs and as a basis for gene editing to generate novel isogenic disease lines

# RBi001-A control line

Test	Result
<b>Derivation centre</b>	R-Biomedical
<b>Disease</b>	Healthy control
<b>Gender</b>	Male
<b>Reprogramming method</b>	Episomal
<b>Passage number</b>	P17
<b>Post-bank Inoculation for microbiological growth.</b>	Not contamination detected
<b>Post-bank Mycoplasma QPCR</b>	Not mycoplasma detected
<b>Viral Screening (HIV1, HIV2, HEP-B and HEP-C)</b>	Negative
<b>Viability post-cryopreservation (post-bank)</b>	Pass
<b>Differentiation Potential</b>	Neuronal lineage confirmed
<b>Phenotype</b>	Normal
<b>Post-bank morphology (EBiSC/FRM/2)</b>	Normal
<b>Post-bank STR</b>	Recorded
<b>Post-bank Flow Cytometry (Roslin Cells)</b>	Pass
<b>STR match between fibroblasts and post-bank thaw.</b>	Pass
<b>Karyology</b>	Normal
<b>Episomal Clearance</b>	Clear

# hiPSC -Underpinning Presumptions



- Cell/tissue functional maturity.

*Embryonic vs adult equivalence?*

- Physiological and disease relevance

*Factor co-dependence? Environment, age?*

- Practical value

*Prospective? Retrospective? Corrective?*

- Cost/benefit value

*Efficacy? Affordability?*



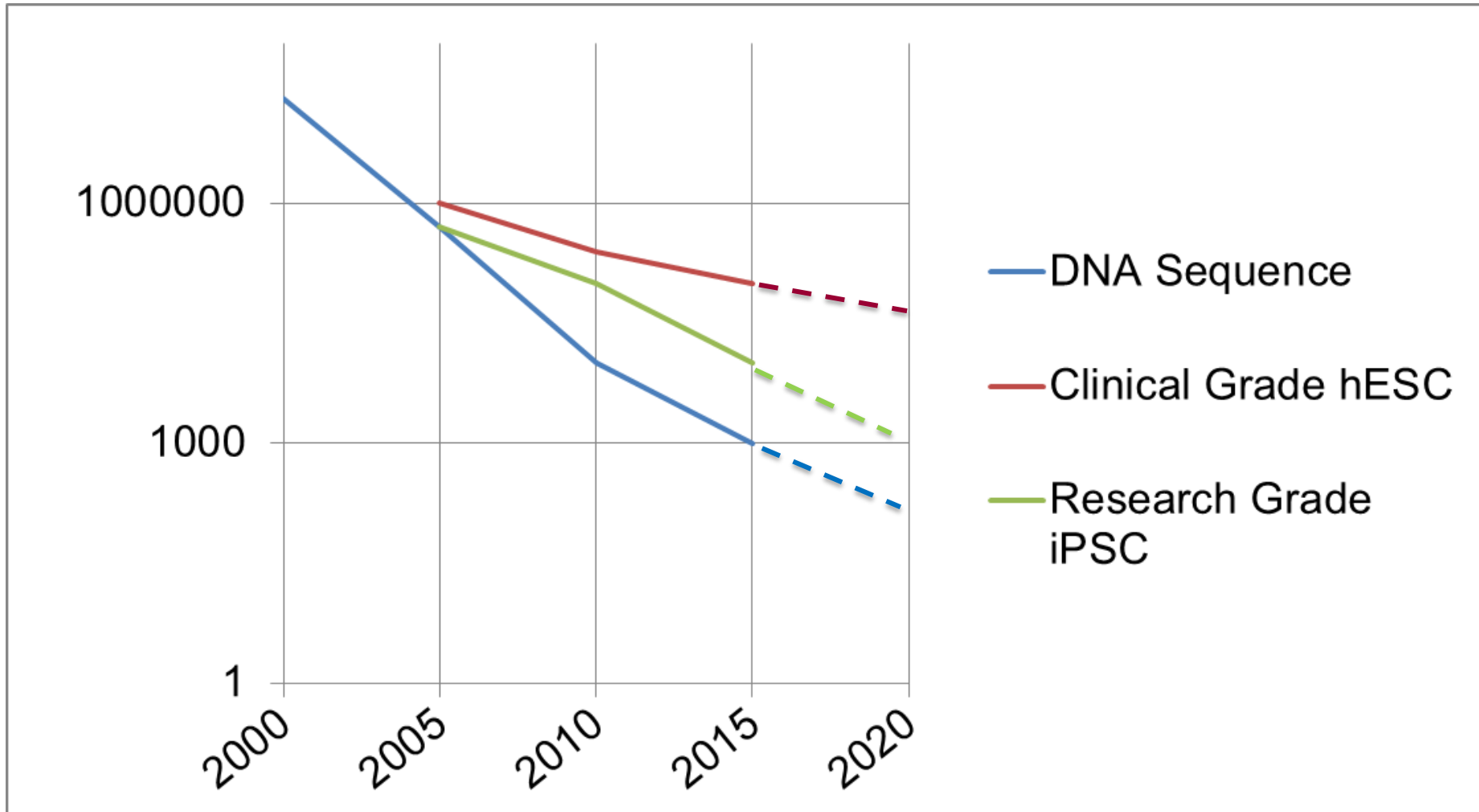
Which of us is representative enough, and when?



*"The People's Monarch"*  
By Helen Marshall  
Pier II Gatwick South Terminal

What do we need to do.....?

# Need to drive cost out of hiPSC production....





# Development of industrialised screening (On the road)



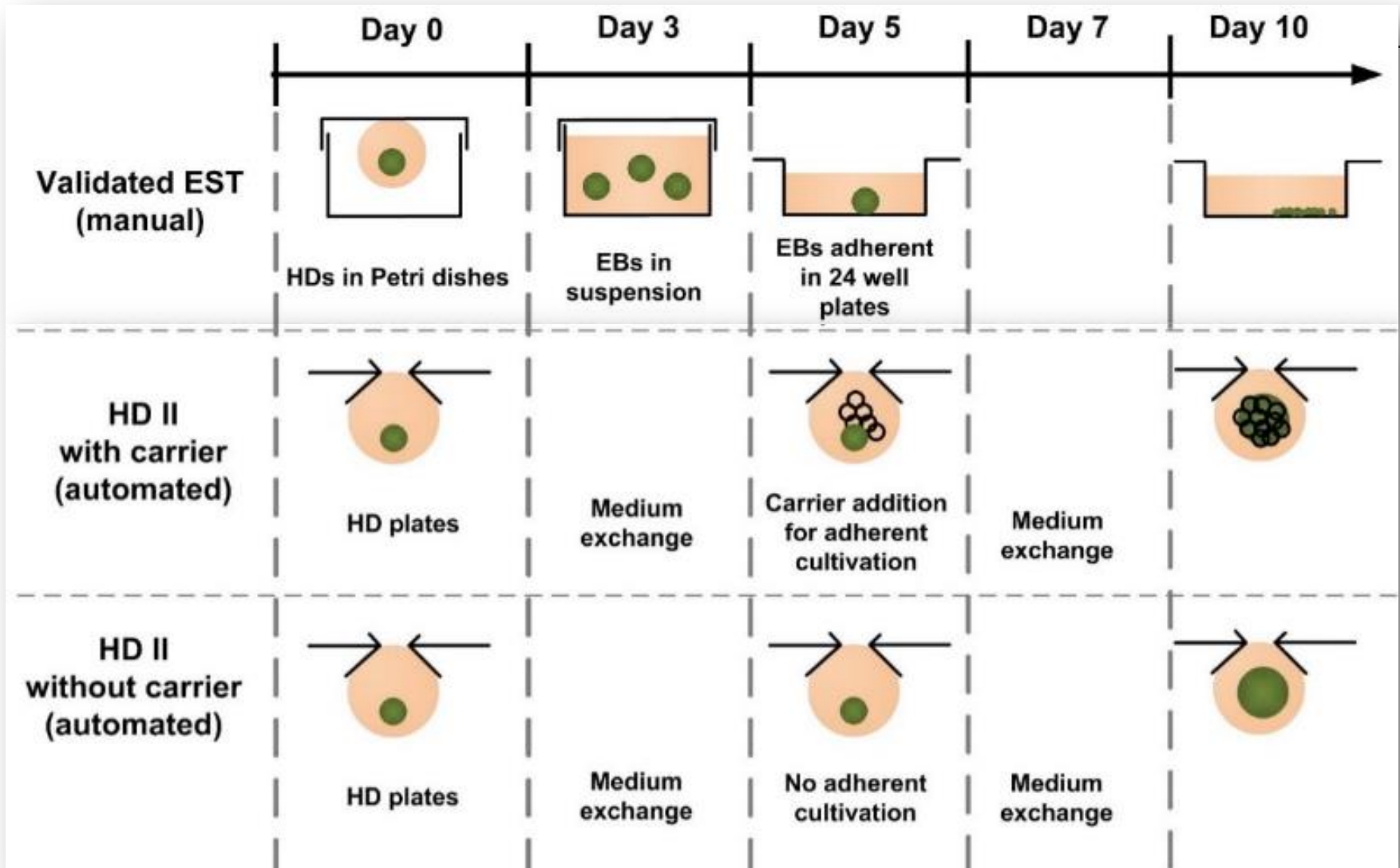
**Aim:** Establishment of an automated hanging drop (HD) human iPSC based version of a mouse ES test (EST) for developmental toxicity.

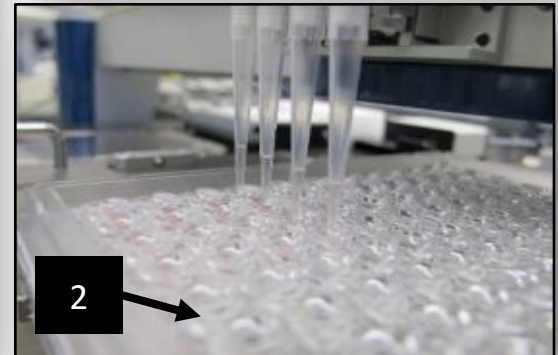
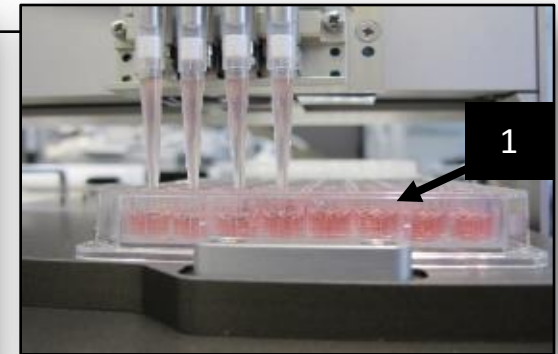
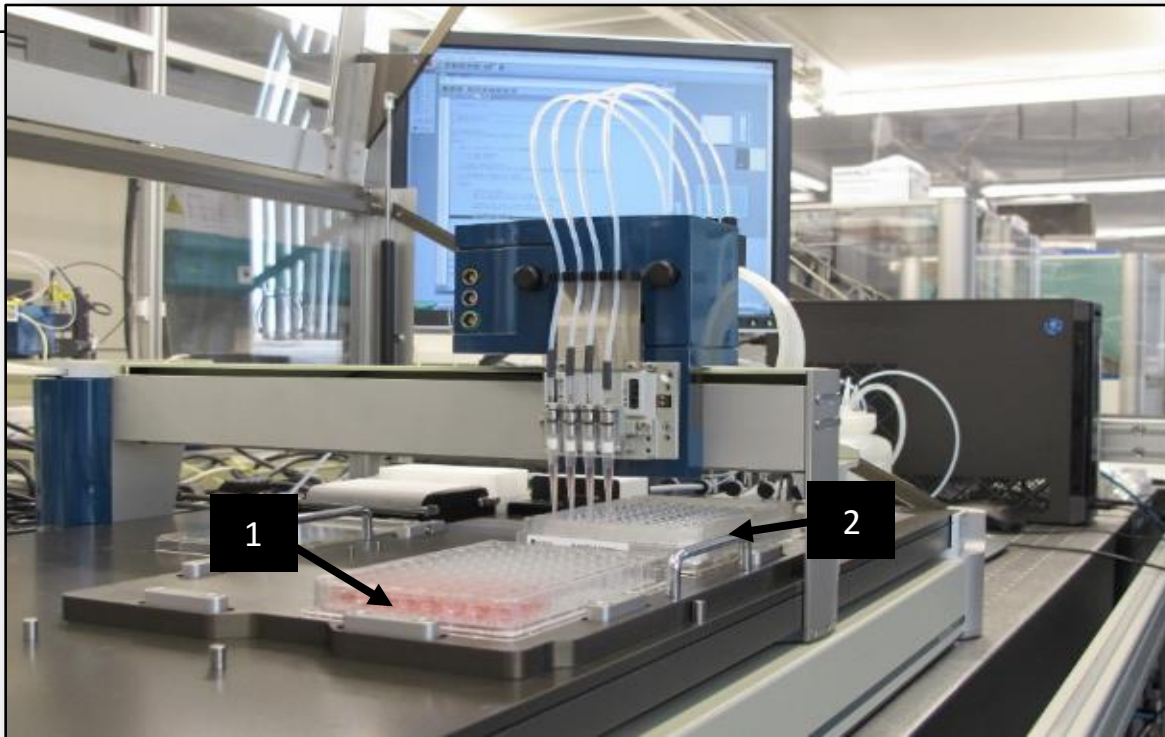
(Seiler & Spielmann 2011, Nature Protocols 6; 961)

**CO:** Prof. Heiko Zimmermann



# Workflow of EST in HDs



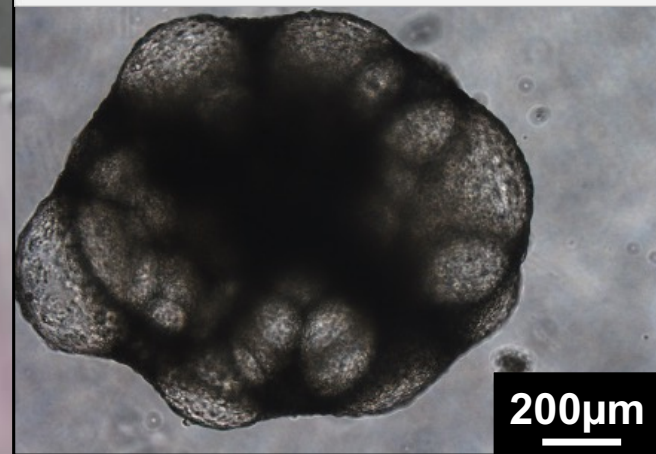
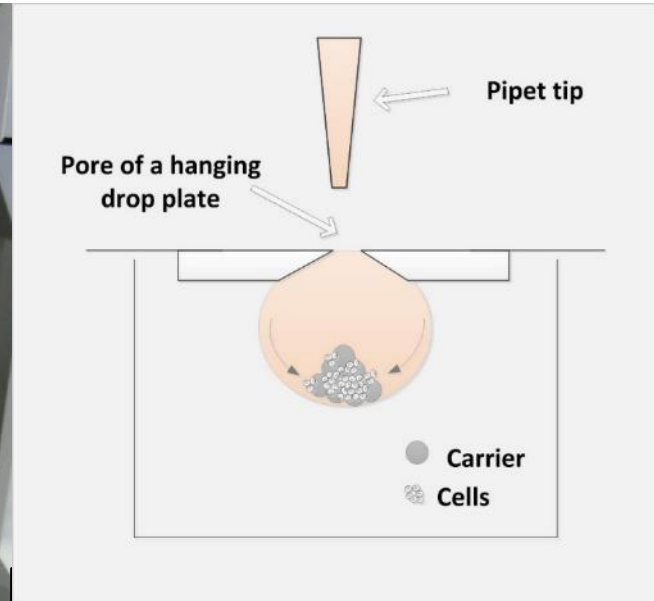


4-channel pipet robot operating an HD plate

(1) Feed plate

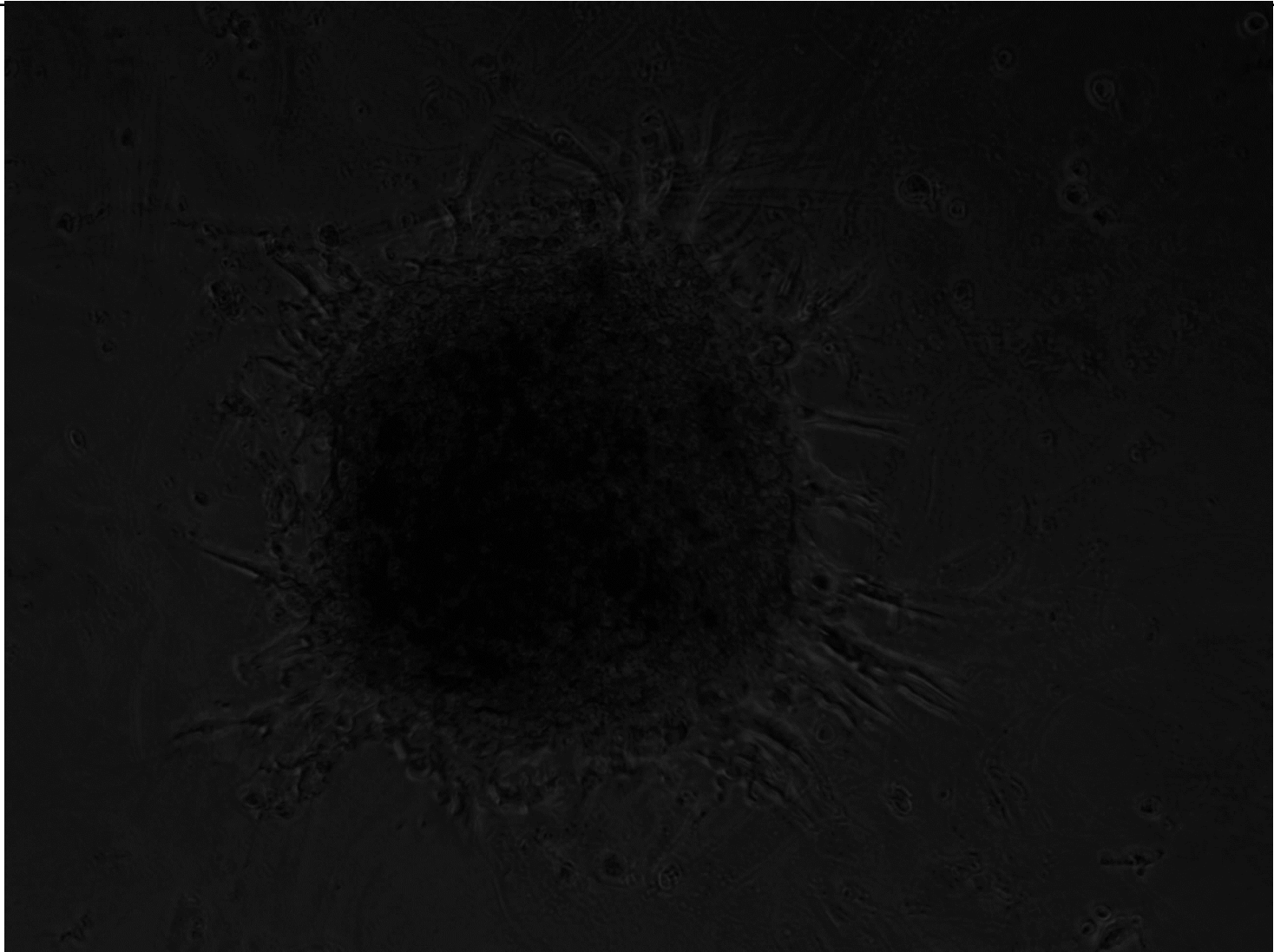
(2) HD plate

# 4-Channel Nanoplotter 2.1 operating Perfecta-3D plates (3D Biomatrix)





# HiPSC derived Cardiomyocyte-like outgrowth



# HD automation with TECAN robot



# What the nice man said.....

- EBiSC standardisation → Industrialisation
- HiPSC lines now available.
- Automated hiPSC HD version of mEST (DROPTTECH).



# Acknowledgements



Droptech Consortium

EBiSC Consortium



